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# **FUTA Journal of Research in** Sciences



ISSN: 2315 - 8239 (Print); E-ISSN: 2489 - 0413

FUTA Journal of Research in Sciences, Vol. 13 (1) 2017:10-16

# Comparative Study on Phytochemical Quantification and Antimicrobial Activity of Raufolvia Vomitoria Leaves, Seeds and Root Extracts

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#### **ABSTRACT**

One of the world medical problems is the challenge of bacteria and fungi resistance to commercial antibiotics. Recently some organisms such as Staphylococcus aureus, Klebsiella pneumoniae, Escherichia coli, and Enterococcus faecalis to mention but a few have developed drug resistance. This study aimed to quantitatively determine the phytochemical constituents of Raufolvia vomitoria seed, leaf and root. It further aimed to evaluate the antimicrobial effect of the ethanolic extract of seed, leaf and root against some selected bacteria (Bacillus subtilis, Bacillus cereus, Pseudomonas syringine, Pseudomonas aeurginosa, Escherichia coli, Xanthomonas axonopodis and Staphylococcus aurus) and fungi (Cyrtomium falcutum, Rhizoctonia solani, Colletotrihum lindematianum, Trichophyton rubrum). Phytochemical analysis was determined using standard procedures and antimicrobial activity was performed by agar well diffusion. The result revealed that the root had the highest alkaloids, phenols, phytates and saponins contents of 15.72 %, 1.24%, 1.66% and 5.99% respectively. The seed had the highest flavonoid contents of 9.38 and the leaf recorded the highest oxalates content of 0.17 %. The result revealed that all the ethanolic extracts are potent against the test microbe. Inhibition zones of the extract against bacteria ranged from 1.20 mm to 10.00 mm while the mycelia inhibition against fungi ranged from 4.76 mm to 26.98 mm. The research demonstrates that extracts from the seed, leaf and root of Raufolvia vomitoria could be used for production of broad pharmaceutical and medicinal based products.

#### Keywords: Rauvolfia vomitoria, phytochemical, bacteria, fungi, Extract and Medicinal

#### INTRODUCTION

One of the world medical challenges is the current trend of multi drug resistance by bacteria and fungi. Recently some organisms such as Staphylococcus aureus, Klebsiella pneumoniae, Escherichia coli, and Enterococcus faecalis to mention but a few have developed drug resistance WHO, (2004) in Niau et al., (2014). Antibiotic resistance has not only increased human mortality rate but also contributed to increase in poverty level, because huge amount of money is being deployed into disease prevention and cure (Njau et al., 2014).

Natural drugs are the early source medicine for human being and extracts from plant's parts (seed, root, flower, stem and bark) have been used for curing various ailments in the time past. Although, the advent of modern medicine has affected the development of local herbal medicine, notwithstanding plant materials are still being used to greater extent for combatting illnesses and various diseases, and researches for novel plant extracts and isolates for the development of new medicine is the focus of many researchers (Ozusaglam and Karakoca, 2013).

Ethnobotanical medicine provides alternative sources of new drugs, an example is Rauvolfia species most found in sub-Sharan Africa and it is an important medicinal plant. They have been effective in the treatment of malaria, diabetes, and both parasitic and microbial infections (Amole et al., 1998; Campbell et al., 2006; Pesewu et al., 2008; Zirihi et al., 2005). In particular, Rauvolfia vomitoria has been used in the treatment of coughs, gastrointestinal disturbances, skin infections, hypertension, impotence, insomnia, diarrhea, dysentery, scabies. worm infections and malaria (Odugbami, 2008; Oyedeji, 2007). The chemical components of this plant include indole alkaloids (Amer and Court, 1980; Katič et al., 1980) . R. vomitoria root contained high concentration of alkaloids; majorly reserpine and ajmaline. These alkaloid's pharmacological abilities include antimalarial, antitumor and antidiabetes (Dewick, 2002; Katič et al., 1980). Reserpine and ajmaline have good antioxidant activities, thus this plant is capable of reducing risks of certain type of degenerative diseases (Erasto et al., 2007; Mazza et al., 1999).

To the best of our knowledge there is no research on the quantitative determination of the root, leave and seed of *Raufolvia vomitoria*, neither is there any reported comparative antimicrobial effect of their ethanolic extracts. The study is therefore aimed at comparing the quantitative phytochemical and antibacterial activity of ethanolic extracts leaves, seeds and root of *Raufolvia vomitoria against* some bacteria and fungi.

#### MATERIALS AND METHODS

#### Plant sample preparation

The seeds, leaves and roots of *R. vomitoria* were obtained from Agbani farm land in Agbado-Ekiti, Gbonyin Local Government Area, Ekiti State, Nigeria. All the samples were authenticated at the Department of Crop, Soil and Pest Management, Federal University of Technology Akure, Ondo State, Nigeria. All samples were cleaned to remove dust, sand, dirt and contaminations. The materials were then dried at  $(27\pm1^{\circ}\text{C})$  room temperature. The samples were pulverized and sieve range from  $212\text{-}249\mu\text{m}$  was used.

# Preparation of extract from seed, root and leaves of *R. vomitoria*

About 50 g plant sample was extracted using 95 % analytical grade ethanol (500 mL) in soxhlet extractor for 8 h. After 8 h the extract was concentrated by exposing the solvent to the air for drying.

#### Quantitative phytochemical analysis

The dried powdered samples of the root, seed and leaf of the plant were subjected to quantitative phytochemical analysis using existing methods and the following were determined; alkaloid (Sathya et al., 2013), total phenol (Hussain et al., 2011), flavonoid (Boham and Kocipai, 1994), phytate (Olajide et al., 2011), oxalate (Oladele et al., 2009) and saponin (Obadoni and Ochuko, 2002).

#### **Antibacterial activities of the extracts**

The antimicrobial activity of the extracts was determined using the agar-well diffusion method of (Murray et al., 2006). 20 mL of nutrient agar was dispersed into sterile petri dish and inoculated. The plates were allowed to set and well of 6mm was properly bored using sterilized cork bore. The wells were filled with the extract of 0.01 mg/ mL. The plates were incubated at 37°C for 24h. The zones of inhibition were measured with Vernier calipers in mm and recorded appropriately. The experiment was done in triplicate and mean values reported. Streptomycin sulphate and ciprofloxacin were used as positive control.

#### Antifungal activities of the extracts

Poisoned food techniques (Mishra et al., 2012) was the method of choice for this investigation, 0.01mg/ml concentration of the samples were prepared and thoroughly mixed separately with 20 ml of potatoes Dextrose agar (PDA) in a sterile sample bottle before pouring in a plate. For the control sets, distill water was used instead of the sample as negative control and 0.01mg/ml ketoconazole was used as positive control. The poured plated were inoculated aseptically with a 6mm fungal disc of 72hrs old of pure culture of the test isolates and incubated between 25-27°C for 72hr for 5days. The observations were recorded at the end of the

incubation period. Percentage mycelial growth inhibition was calculated by following formula; Percentage growth inhibition =  $\frac{NTR-TR}{NTR}$  X 100

Where, NTR= average diameter of fungal colony in negative control set (without treatments)

TR= average diameter of fungal colony in treatment sets

The experiments were carried out in triplicate and the average values recorded was as shown in the table below

#### **Statistical Analysis**

The data obtained in triplicate were analysis by Probit Analysis using Duncan's Multiple Range Test (DMRT) and Analysis of Variance (ANOVA)

#### RESULTS AND DISCUSSION

Preliminary phytochemical screening of the leaf, stem and root of Rauwolfia vomitoria according to (Adebayo et al., 2014) revealed the presence of alkaloid, phenols and flavonoids, The results of the quantitative phytochemical analysis of the root, leaf and seed of Rauwolfia vomitoria are presented in Table 1. The analysis reveals the presence of alkaloids, flavonoids, phenols, phytates, oxalates and saponins to different degree. The high alkaloids content (15.72 %) in the root was in accordance with findings of others, that had found that the root has high alkaloid content (Dewick, 2002; Katič et al., 1980). The seed has 14.90 % and leaf has 3.16 % alkaloids content. The high alkaloids content of the root and seed might be because those parts serve have storage faculty of the plant (Benbott et al., 2012).

whereas (Ojo et al., 2012), presented the alkaloid content (2.21, 2.23 and 3.39 %), flavonoids (0.37, 0.39 and 2.18 %), oxalates (3.69, 3.78 and 3.06 mg/g) and polyphenols (2.10, 1.91 and 1.35) respectively in aqueous, ethanol and methanol extracts of the back of *Rauwolfia vomitoria*. The results of these authors confirmed the presence of various phytochemicals determined in this study, although different results were obtained. This

was expected because of the plant materials were not obtained at the same place, seasonal variation could also affect the result likewise some laboratory experimental errors.

In our study the flavonoids contents are 9.38 % (seed), 8.60 % (leaf) 7.56 % (root), showing that the seed has highest content and this may be attributed to the reddish color of the seed. The root had the highest phenols 1.24 %, while 0.32 % and 0.08 % were recorded for the leaf and seed. The high phenolic content of the root might be because they act as pest and disease resistance agent for the effective growth of the plant (Li et al., 2010).

Phytates salts of phytic acid, have been implicated to reduced absorption of dietary minerals both in human and animal (Konietzny and Greiner, 2003; Lopez et al., 2002). Consumption of phytates rich food, however has greater health benefit than the negative effect of reduced dietary mineral absorption. Dietary phytates has been reported to have medicinal value to prevent and protect kidney stone formation (Grases et al., 2000), atheriosclerosis and coronary heart disease (Jariwalla et al., 1990) and a variety of cancers (Vucenik and Shamsuddin, 2003). Therefore, the moderate phytates recorded in this research 1.66 %, 1.52 % and 0.48 % respectively for the root, leaf and seed would be of health benefit to both human and animals for phytates supplement. The percentage oxalates recorded the lowest value of the phytochemical. The seed had 0.00 %, root has 0.08 % and the leaf 0.17 %. The results seem encouraging knowing fully well that the plant is being used for various medicinal purposes and the low oxalates content would reduce the risk of calcium oxalate urinary stone formation. The root contained the highest percentage (5.99 %) of saponins, followed by the leaf (4.80 %) and then the seed (4.56 %).

The antimicrobial results of the crude ethanolic extracts of the root, seed and leaf *R. vomitoria* are as presented in Table 2 and 3 at 0.01 mg/mL extract and control concentration. As presented in Table 1 the determined chemical components of the plant suggest presupposes antibacterial activities of the plant parts. The result indicated that those extracts were active against the entire microbes with zones of inhibition that varied from 4.76 to 26.56 mm for fungi. The antimicrobial results proof the potency of the extracts with broad-antibacterial activities as

they are active against both Gram-positive and Gram-negative bacteria. Their extracts are active against pathogenic bacteria such as *E. coli, Bacillus cereus, B. subtilis, S.aureus, X. axonopodis, P.syringine* and *P.aeuru* with zone

of inhibition ranging from 1.50 to 10.00 mm. Although, all the control used were much more active than the crude plant extracts, the result shows that the fungi were more susceptible to the plant extracts than the bacteria

Table 1- Quantitative Phytochemical of Raufolvia vomitoria

Values are means of three replicate  $\pm$  standard deviation. Column means followed by different letters are significantly different at P< 0.05

Table 2- Zones of inhibition (mm) of the extracts obtained from root, seed and leaf of R. vomitoria at 0.01 mg/mL

Sample	B.subtilis	X.axonopodis	B.cereus	P.syringine	P.aeruginosa	E.coli	S.
							aureus
RVS	3.00	4.00	6.00	4.00	5.00	3.00	7.00
RVL	2.00	7.00	5.50	10.00	3.25	7.50	8.00
RVR	1.50	3.00	3.50	3.00	1.45	1.20	4.00
Streptomycin	26.00	11.50	15.00	14.50	15.00	15.00	16.00
Ciprofloxacin	17.00	12.00	15.00	28.00	20.00	26.00	24.00

Where, RVS is *Raufolvia vomitoria* seed, RVL *Raufolvia vomitoria* leaf and RVR is *Raufolvia vomitoria* root

Table 3- Percentage growth inhibition of extracts obtained from root, seed and leaf of R.  $vomitoria\ at\ 0.01\ mg/mL$ 

Sample	C.falcutum	R. solani	C. linalimutianum	T. rubrum
RVS	4.76	17.95	20.15	18.75
RVL	26.98	9.62	34.33	21.56
RVR	25.40	17.30	13.73	26.56
Ketoco	60.00	85.00	53.73	88.88

Where, RVS is Raufolvia vomitoria seed, RVL Raufolvia vomitoria leaf and RVR is Raufolvia vomitoria root

Ketoco= Ketoconazole (a standard antifungal agent)

The high percentages of alkaloids, saponins, and flavonoids in the root, seed and leaf of R. *vomitoria and* moderate presence of phytates, phenols and oxalates could make the plant a good source of ethnobotanical medicine.

According to Vignesh and Chandra, (2015), alkaloid in *Rauvolfia canescens* leaf was attributed to its antibacterial potency, while Sonibare et al., (2011) revealed that alkaloids and saponins in *Rauvolfia vomitoria* leaf

conferred on it antimicrobial activities. Also, Mert-Türk, (2006) stated that phytates, phenols saponins and alkaloids and some other secondary metabolites in plants play important role in plant defense against pathogens. The natural protection role of these phytochemicals against plant pathogens could be harnessed for medicinal and pharmaceutical benefit of human being. The results obtained in this study were in agreement with antimicrobial potency of plant materials using Rauvolfia vomitoria leaf (Sonibare et al., 2011), Rauvolfia canescens leaf (Vignesh and Chandra, 2015) and using stem, leaf and root extracts of Rauvolfia vomitoria against some pathogens, although with the use of different pathogen (Adebayo et al., 2014).

#### **CONCLUSION**

The broad antifungal and antibacterial activities of the plant could be linked to the presence of phytochemicals in the plant. Therefore, in the face of microbes resistances to modern drugs, isolation, purification and chemical functionalization of phytochemical presence in this plant could be a good solution to the failure witnessed in some drugs during diseases treatment.

#### REFERENCES

### Adebayo, O. L. Unique A. B. M. and

**Sunyazi S. S.** (2014). In vitro bacteria growth inhibition potential of crude extracts of different parts of Rauvolfia vomitoria against five standard strains of selected human pathogenic microbes. World Journal of Pharmacy and Pharmaceutical Sciences, 3(6), 152-164

Amer, M. M. and Court W. E. (1980). Leaf alkaloids of Rauwolfia vomitoria, Phytochemistry. 19 (8), 1833–1836.

Amole, O. O. Onabanjo A. C. Agbaje E. C. (1998). Effects of bark extracts of Rauvolfia vomitoria (afzel) in malaria. Parasitology International. 47, (1), 380.

Benbott, O. O. Yahyia A. and Belaidi A. (2012) . Assessment of the Antibacterial Activity Crude Alkaloids Extracted from Seeds and Roots of the Plant Peganum

harmala. Journal of Natural Products and Plant Resources, 2(5), 568-573.

## Boham, A. and Kocipai A. (1994).

Flavonoid and condensed tannins from. Leaves of Hawaiian vaccininum vaticulum and vicalycinium. Pacific Science. 48, 458–463.

# Campbell, J. I. A. Mortensen A. and

Mølgaard P. (2006). Tissue lipid lowering-effect of a traditional Nigerian anti-diabetic infusion of Rauwolfia vomitoria foilage and Citrus aurantium fruit. Journal of Ethnopharmacology. 104, 379–386.

**Dewick, P. M.** (2002). Medicinal Natural Products A biosynthetic approach, 2nd Ed. ed. John Wiley and Sons Ltd.

Erasto, P. Grierson, D. and Afolayan, A. (2007). Evaluation of Antioxidant activity and the fatty acid profile of the leaves of Vernonia amygdalina growing in South Africa. Food Chem. 104, 636–642.

Grases, F. March J. G. Prieto R. M. Simonet B. M. Costa-Bauzá A. García-Raja A. and Cont A. (2000). Urinary phytate in calcium oxalate stone formers and healthy people--dietary effects on phytate excretion. Scandinavian Journal of Urology and Nephrology. 34, 162–164.

# Hussain, I. Ullah R. Ullah R. Khurram M. Ullah N. Baseer A. Khan F. Rehman M. (2011). Phytochemical analysis of selected medicinal plants. African Journal of Biotechnology. 10, 7487–7442.

**Jariwalla, R. J. Sabin R. Lawson S. and Herman Z. S.** (1990). Lowering of serum cholesterol and triglycerides and modulation of divalent cations by dietary phytate. Journal of Applied Nutrition. 42, 18–28.

- **M.** (1980). Quantitative densitometric determination of reserpine and ajmaline in Rauwolfia Vomitoria by HPTLC. Journal of High Resolution Chromatography. 3, 149–150.
- Konietzny, U. and Greiner R.(2003).

  Phytic acid: Nutrition Impact. In;
  Encyclopedia of Food Science and
  Nutrition, B. Caballero, L. Trugo, P.
  Finglas Eds. ed. Elsevier, London, UK.
- **Li, Z.-H. Wang Q. Ruan X. Pann C.-D.** and Jiang D.A. (2010). Phenolics and Plant Allelopathy. Molecules 15, 8933–8952.
- Lopez, H. Leenhardt F. Coudray C. and Remesy C. (2002). Marschner's Mineral Nutrition of Higher Plants. International Journal of Food Science & Technology. 37, 727–739.
- Mazza, G. Fukumoto L. Delaquis P. Girard B. Ewert B. (1999). Anthocyanins, phenolics, and color of Cabernet Franc, Merlot, and Pinot Noir wines from British Columbia. Journal of Food Chemistry. 47, 4009–4017.
- **Mert-Türk, F.** (2006). Saponins versus plant fungal pathogens. Journal of Cell and Molecular Biology. 5, 13–17.
- Mishra, P. K. Shukla R. Singh P.

  Prakash B. Kedia A. and Dubey N.

  K. (2012). Antifungal, antiaflatoxigenic, and antioxidant efficacy of Jamrosa essential oil for preservation of herbal raw materials. International Biodeterioration & Biodegradation. 74, 11–16.
- Murray, P. Rosenthal K. and Pfaller M. (2006). Medical Microbiology, Fifth Edition. ed. Spain. Elsevier.
- Njau, E. F. Alcorn J. Chirino-Trejo P. N. and Buza, J. (2014). Antimicrobial and Antioxidant Activity of Crude Extracts of Rauvolfia caffra var. caffra

- (Apocynaceae) from Tanzania. International Journal of Biology. 6, 156–1667.
- Obadoni, B. and Ochuko P. (2002).

Phytochemical Studies And Comparative Efficacy of the Crude Extracts of Some Haemostatic Plants In Edo And Delta States Of Nigeria. Global Journal of Pure and Applied Sciences. 8, 203–208.

- Odugbami, T. (2008). A Textbook of Medicinal Plants from Nigeria. University of Lagos Press, Lagos, Nigeria.
- Ojo, O. Ajayi S. and Owolabi, L. (2012).

Phytochemical screening, anti-nutrient composition, proximate analyses and the antimicrobial activities of the aqueous and organic extracts of bark of Rauvolfia vomitoriaand leaves of Peperomia pellucida. International Research Journal of Biochemistry and Bioinformatics. 2, 127–134.

#### Oladele, K. Osundahunsi F. and

**Adebowale A.** (2009). Influence of Processing Techniques on the Nutrients and Antinutrients of Tigernut (Cyperus esculentus L.). World Journal of Dairy and Food Sciences. 4, 88–93.

- Olajide, R., Akinsoyinu, A. O. Babayemi
  O. J. Omojola A. B. Abu A. O.
  Afolabi K. D. (2011). Effect of
  Processing on Energy Values, Nutrient
  and Anti-nutrient Components of Wild
  Cocoyam [Colocasia esculenta (L.)
  Schott] Corm. Pakistan Journal of
  Nutrition. 10, 29–34.
- **Oyedeji, L.** (2007). Drugless Healing Secret. Ibadan, Panse press.
- Ozusaglam, M. A., Karakoca K. (2013)
  Antimicrobial and antioxidant activities of *Momordica charantia* from Turkey .
  African Journal of Biotechnology. 12 (13): 1548-1558
- Pesewu, G. A. Cutler R. R. Humber D.

- **P.** (2008). Antibacterial activity of plants used in traditional medicines of Ghana with particular reference to MRSA. Journal of Ethnopharmacology. 116(1), 102-111.
- Sathya, V. Bharathidasan R. Tamiln S. Sophia R.nIlakkiya R. Prabakaran, M. (2013). Quantitative, qualitative phytochemical analysis and vitro anti bacterial activity of Bauhinia tomentosa L. Journal of Natural Product and Plant Resources. 3, 31–36.
- Sonibare, M. A. Lawal T. O. and
  Ayodeji O. O. (2011). Antimicrobial
  Evaluation of Plants Commonly Used in
  the Management of Psychosis
  Opportunistic Infections. International
  Journal of Pharmacology. 7, 492–497.

**Vignesh, T. and Chandra, J. H.** (2015). Antibacterial activity and phytochemical analysis of leaf extracts of Rauvolfia

canescens. Journal of Chemical and Pharmaceutical Research. 7(4), 11-14

- Vucenik, I. and Shamsuddin, A. M. (2003). Cancer Inhibition by Inositol Hexaphosphate (IP6) and Inositol: From Laboratory to Clinic. Journal of Nutrition. 133, 3778S–3784S.
- **Zirihi, G. N. Mambu, L. Guédé-Guina F. Bodo, B. Grellier P.** (2005). In vitro antiplasmodial activity and cytotoxicity of 33 West African plants used for treatment of malaria. Journal of Ethnopharmacology. 98, 281–285.