



## Effect of combined levels of *Panicum maximum* and *Gliricidia sepium* on the anti-Nutrients factors' digestibility in West African Dwarf Does

<sup>1</sup>OGUNGBESAN, A.M., <sup>1</sup>FASINA, O.E., <sup>2</sup>AYUK, A.A. and <sup>3</sup>LAMIDI, A.

<sup>1</sup>Department of Animal Production, Olabisi Onabanjo University, Ayetoro, Ogun State, Nigeria.

<sup>2</sup>Department of Animal Science, University of Calabar, Cross River State, Nigeria.

<sup>3</sup>Department of Animal and Fisheries Sciences, University of Port-Harcourt, Rivers State Nigeria.

**ABSTRACT:** Thirty (30) West African Dwarf does of weaning weight of  $5.86 \pm 1.20$ kg, balanced for weight, and age were used to investigate the effect of combinations of *Panicum maximum* (Pm) and *Gliricidia sepium* (Gs) with cassava offal basal diets on antinutrients' digestibility. The five (5) treatments were I (Go) 100% Pm + 0% Gs, II (G<sub>25</sub>): 75% Pm + 25%Gs, III (G<sub>50</sub>): 50% Pm + 50% Gs, IV (G<sub>75</sub>): 25% Pm +75% Gs, V (G<sub>100</sub>): 0% Pm + 100% Gs, Highest (P < 0.05) (75.00%) and lowest (55.80%) digestible oxalate were recorded in G<sub>100</sub> and G<sub>75</sub> respectively. Highest digestible phytate (76.44 %), Tannin (77.17%) Saponin (75.00%) and lowest (P < 0.05) Phytate (57.35%), Tannin (60.70%), and Saponin (58.33%) were observed in Go, G<sub>100</sub>, Go with G<sub>100</sub> and G<sub>75</sub> respectively while the cyanide content was completely digested in all the treatments. The results were not easily attributable to various combinations but it could be seen that goats can tolerate ANF in both sole grass and legume as well as basal diet effectively.

**Keywords:** Antinutritional factor, digestibility, *Panicum*, *Gliricidia*, West African Dwarf Does

JoST. 2014. 5(1): 93-100.

Accepted for Publication, August 25, 2014

### INTRODUCTION

The importance of animal protein in human diet cannot be overemphasized, also other products (wool, skin, horns, fat, hormones/biologics, bone, milk, etc.) play important role in the sustainability of mankind (Wilson, 1995). Goats have survived, produced and reproduced due to selective intake of wide varieties of trees, forbs, grasses, herbs and non-convention feed stuff due to the fact that they are browse-preferring intermediate feeders which exhibit high dietary plasticity there by enabling them to switch feed groups when preferred ones are in limited supply (Strangel, 1993). Traditionally, the off-take (extraction) rate of males (bucks) are more than the females (does) because of

reasons like zero-prolificacy, sex ratio (1 male to 25 females) ritual purposes, festivity, and unprecedented sales to meet petty contingent expenditure (Devendra and Burns, 1985). Guinea grass (*Panicum maximum*) is a widely distributed tussock, lump-forming grass that grows best on warm frost-free areas. It is considered the best grass for ruminant production; it is highly relished by goats, readily available and stimulates ruminal microbial growth (Anugwa *et al.*, 2000). The existence of multipurpose tree legume like *Gliricidia* (*Gliricidia sepium*) which is perennial in nature, has high crude protein and also has been identified as one of the fodder legumes that promotes rumen ammonia

\*Correspondence to: Ogungbesan, A.M., amkgbesan@gmail.com

production and live weight gain (Ajayi *et al.*, 2005). The search for alternative feed resources has focused attention on cassava and its by-products such as cassava seviate, offal and peelings. These by-products are available all year round especially now that the economy is being redirected towards non-oil products. The evolution of these by-products which are not directly useful to man can be channeled towards the production of animals and their products thereby reducing over dependence on cassava root tubers. Also, the adverse environmental pollution caused by these by-products can be reduced. The afore-mentioned feeds contain some inherent antinutritional factors as catalogued by Norton (1994) which include oxalate, tannin, cyanogenic glucosides, flavonoids, coumarin, just to mention a few. Researchers (Onwuka, 1990, Oduguwa *et al.*, 1998, and Ogungbesan *et al.*, 2005, Ogungbesan

*et al.*, 2006) have shown that some of these could be tolerated by ruminants because ruminal microbes secrete enzymes that can degrade these deleterious principles. In addition to this, some plants have inherent “antinutritional factors that hydrolyse these substances (Eeckhoat and Depaepe, 1994) Works have also been done on Gliricidia and Guinea grass on goats (Smith *et al.*, 1995, Ajayi *et al.*, 2005,) while this combinations had been fed with cassava and Brewers dried grain BDG. (Ifut, 1992). More recently Eniolorunda *et al.*, (2008) researched into the nutrients digestibility and utilization of the aforementioned feed stuff but there is no information on the anti nutrient digestibility in animals fed these combinations. This study therefore investigated the effect of combined levels of *Panicum maximum* and *Gliricidia sepium* on the antinutrients factors' digestibility in W.A.D goats.

#### MATERIALS AND METHODS

The study was conducted at the goat unit of Olabisi Onabanjo University, Yewa Campus, Ayetoro, Ogun State, Nigeria. Ayetoro is located on latitude 7°15'N and longitude 3°3'E in a deciduous derived savannah zone of Ogun State. The climate is sub-humid tropic with an annual rainfall of 963.3mm in 74 days. Temperature varies between 29 and 34°C.

Thirty growing Wet African Dwarf does aged 5-7 months and with an average live weight of 5.86kg were used for this experiment. They were obtained from donor farms located some 15km northwest of the experimental site. Two weeks before the commencement of the experiment, animals were dewormed with Levamisole (Kepro, B.V Holland, 1mL/per 20kg body weight) to control endoparasites and dipped in Diazintol solution (Alfasan International, B.V Holland) at the rate of 1mL per liter of water against ectoparasites. Long acting Oxytetraciclina 200 La (Invesa, Spain) at 1 mL per 10kg body weight

was also administered. The animals were randomly allocated on live weight basis to 5 groups of 6 does each. The 5 groups were then moved into previously sanitized individual pens and offered the experimental diets daily for 98 days (including the first 14days for adaptation and subsequent 84days for measurement).

The dietary treatments were:

- 100% *Panicum maximum* and 0% *Gliricidia sepium* (G 0).
- 75% *Panicum maximum* and 25% *Gliricidia sepium* (G 25).
- 50% *Panicum maximum* and 50% *Gliricidia sepium* (G 50).
- 25% *Panicum maximum* and 75% *Gliricidia sepium* (G 75).
- 0% *Panicum maximum* and 100% *Gliricidia sepium* (G 100).

Combined levels of *P. maximum* and *G. sepium* were supplemented with a cassava offal-based concentrate. The *G. sepium* foliage were allowed

to wilt overnight prior to feeding. *P. maximum* leaves were harvested daily, manually chopped to 5cm and fed to the animals fresh. All animals were fed twice daily (forages at 3% body weight by 08.00h and concentrate by 16.00h at 2% BW) and fresh water was available at all times. Cassava offal used for the study was collected from four Fufu processing centers in Ayetoro, Ogun State and dried to a moisture content level of about 12% before being used to compound a concentrate ration containing 16% CP and 2600Kcal kg<sup>-1</sup> Me. The concentrate included 40% cassava offal, 15% cassava peels, 15% GNC, 25% PKC, 4% bone meal, 0.5% mineral premix and 0.5% salt. The animals were weighed once weekly and the level of feeding adjusted depending on liveweight changes. Daily voluntary intake was estimated by difference of the feed offered and residue collected. Digestion and nitrogen balance trial was conducted after 72 days of feeding. During the metabolism trial, the goats were housed in individual metabolism cages (90cm x 75cm x 90cm) made of welded wire mesh fitted with removable feeders and arranged for quantitative collection of faeces and urine separately. The trial lasted for 12 days with a five-day adaptation period to accustom the goats to cages prior to a 7 days collection and measurement period. Total faecal output and urine were collected in the morning before feeding and watering. The faeces were weighed fresh and 10% aliquots of each day's collection for each animal were taken, dried at 60°C for 48h in a forced draught air oven and bulked. A sub sample from each animal was dried in a similar oven at 100-105°C for 48h for dry matter (DM)

determination. Another sub-sample was thoroughly mixed and milled to pass through a 0.60mm sieve and stored in sealed polythene bags until analysed. The urine was collected in a plastic tray placed under each cage. 10mL of 10% concentrated H<sub>2</sub>SO<sub>4</sub> was added to the tray daily to prevent volatilization of NH<sub>3</sub> from the urine. The total output of urine per animal was measured and 10% aliquots were kept in stoppered numbered plastic bottles and stored at -5°C. Methods employed for feed and faecal samples were as follows: Phytate (Maga, 1983), Oxalate (Beutler, et al., 1980) Saponin (Strong, 1976), Tannin (Hagerman and Butler, 1983) and cyanide (Poonam and Hahn, 1984).

#### Statistical Analysis

Data obtained from these chemo-assays were used to calculate the metabolites' digestibility and were further subjected to analyses using one-way ANOVA/completely randomized design using individual goat as replicates. Model sums of square were partitioned to test linear and quadratic trends of supplementation using the General Linear models (GLM) procedures SAS (2002) and significantly different means were separated using least significant difference at 0.5 level of probability.

The general linear model is defined thus

$$X_{ij} = \mu + \alpha_i + e_{ij}$$

$\mu$  is Grand population mean

$X_{ij}$  is individual data generated from the fixed treatment effects

$\alpha_i$  is the fixed treatments (diets I to V) effects

$e_{ij}$  is the error (replicate) term within each treatment

### RESULTS AND DISCUSSION

Table 1 shows the chemo-metric of the anti-nutritional factors analysed for in respective feed components. These from nutritional stand point could be undesirable to some extent but ensure the coevolution of these herbs with herbivores

(Rosenthal and Janzen, 1979), They (i.e. tannin and saponin) at times in small quantity are desirable and advantageous to the animals consuming them. Oxalate was not detected in *P. maximum*. This is contrary to the findings of

**Table 1: Chemo-metrics of deleterious principles in the feed components (mg/100g)**

| Composition | <i>P. maximum</i> | <i>G. sepium</i> | Concentrate |
|-------------|-------------------|------------------|-------------|
| Oxalate     | ND                | 1.25             | 1.16        |
| Phytate     | 98.45             | 101.22           | 113.51      |
| Cyanide     | ND                | 0.15             | 0.34        |
| Tannin      | ND                | 2.65             | 3.06        |
| Saponin     | ND                | 0.10             | 0.11        |

ND: Not Detected

Skerman and Riveros (1990) and Esminger and Olentine (1980) who established that tropical grasses contain Oxalate even more than any other herbage but the findings of Ogungbesan *et al.*, (2006) is in consonance with the works of the above two authors who recorded 8.16% DM oxalate in Guinea grass.

However, all the previous workers failed to mention whether their oxalate was in soluble or insoluble form (Tangendjaja and Wina, 1995), Oxalate recorded in *Gliricidia sepium* was more than those documented by Ologhobo (1989) in browses (0.52-0.82mg/100gm) and less than that catalogued by Aletor and Omodara (1994) of *Gliricidia sepium* (222.41mg/100gm) and *Leucaena leucocephala* (88.16mg/100gm). Phytate (mg/100g) content unusually observed in *P. maximum* was 94.45 while that in *G. sepium* (101.22) was higher than that of *G. sepium* (16.18mg/100gm) recorded by Aletor and Omodara (1994) as well as higher or lower than those browses recorded by these authors. The cyanide content (mg/100gm) of *G. sepium* is 0.15, although its presence may seem erratic or unusual, it could be Holocalin, p=Glucosyl-oxyman deloritrile, pro-acacipetalin, prunasin, sambunigrin, or vicianin which are cyanides present in leguminous plants (Conn. 1981). The tannin (mg/100gm) of the legume (2.65) was at variance with the findings of Budi and Wina (1995) who did not detect any property of tannin activity in *G. sepium*. Its saponin (mg/100gm) was 0.10 and did not also conform with that

reported by Onwuka (1992) who obtained 0.33% DM saponin in *G. sepium*. All the antinutrients factors detected in the concentrate might have been from various ingredients of the ration. Various factors are however, responsible for variabilities in chemical composition of herbages. These include plant parts, age, season, soil fertility, species, cultivar, post harvest treatments, growing conditions (water stress, drought stress, photo-periodicity) (Rosenthal and Janzen, 1979).

In Table 2, the apparent digestibility of the deleterious principles is shown. The term apparent was added because the digestibility did not anticipate their subsequent absorption in the gastro intestinal tract, but their degradation or destruction enzymatically in the rumen. The intake of all these secondary plant metabolites might have been occasioned by those present in the various combinations of feed consumed. The highest and lowest (P<005) oxalate digestibility was observed in G100 and G25 which implies that the levels have little or no effect on the degradation pattern though there was degradation of at least more than half of those ingested by the rumen microbes (Preston and Leng, 1987). Oxalate complexes calcium, phosphorus and decreases protein availability but does not chelate zinc. Although Phytate is often used interchangeably in literature with Phytic acid and Phytin, Phytic acid, phytate and phytin refer respectively to the free acid, the salt, and the calcium and magnesium salt (Evers, *et*

Table 2: Antinutritional factors' Intake and digestibility in West African Dwarf Does fed various combinations of Grass/Legume with cassava offal based diet

| Parameters             | Treatments          |                     |                     |                    |                    | SEM Probability <sup>b</sup> |     |    |
|------------------------|---------------------|---------------------|---------------------|--------------------|--------------------|------------------------------|-----|----|
|                        | G (control)         | G25                 | G50                 | G75                | G100               |                              | L   | Q  |
| Feed Oxalate (g/d)     | 0.93 <sup>e</sup>   | 1.75 <sup>d</sup>   | 2.45 <sup>c</sup>   | 2.90 <sup>b</sup>  | 3.78 <sup>a</sup>  | 0.15                         | XX  | X  |
| Faecal oxalate (g/d)   | 0.25                | 0.63                | 0.76                | 1.28               | 0.98               |                              |     |    |
| Digestible oxalate (%) | 73.16 <sup>a</sup>  | 59.24 <sup>b</sup>  | 60.09 <sup>b</sup>  | 55.80 <sup>c</sup> | 75.00 <sup>a</sup> | 12.61                        | NS  | XX |
| Feed phytate           | 115.73 <sup>a</sup> | 107.79 <sup>b</sup> | 102.76 <sup>c</sup> | 91.76 <sup>d</sup> | 93.04 <sup>d</sup> | 20.41                        | XX  | NS |
| Faecal phytate         | 27.17               | 32.95               | 43.67               | 39.14              | 24.90              |                              |     |    |
| Digestible phytate (%) | 76.42 <sup>a</sup>  | 69.43 <sup>c</sup>  | 57.50 <sup>c</sup>  | 57.35 <sup>c</sup> | 73.52 <sup>b</sup> | 10.00                        | X   | XX |
| Feed cyanide (g/d)     | 0.32 <sup>c</sup>   | 0.37 <sup>d</sup>   | 0.47 <sup>c</sup>   | 0.61 <sup>b</sup>  | 0.74 <sup>a</sup>  | 0.09                         | Xxx | NS |
| Faecal cyanide (g/d)   | 0.00                | 0.00                | 0.00                | 0.00               | 0.00               |                              |     |    |
| Digestible cyanide (%) | 100                 | 100                 | 100                 | 100                | 100                | 0.00                         | NS  | NS |
| Feed Tannin (g/d)      | 1.93 <sup>d</sup>   | 2.87 <sup>c</sup>   | 4.30 <sup>b</sup>   | 4.97 <sup>b</sup>  | 6.34 <sup>a</sup>  | 0.69                         | XXX | NS |
| Faecal Tannin (g/d)    | 0.54                | 0.93                | 1.39                | 1.95               | 1.44               |                              |     |    |
| Digestible Tannin (%)  | 72.20 <sup>b</sup>  | 67.11 <sup>c</sup>  | 67.67 <sup>c</sup>  | 60.70 <sup>d</sup> | 77.17 <sup>a</sup> | 8.52                         | XX  | NS |
| Feed Saponin (g/d)     | 0.25 <sup>c</sup>   | 0.40 <sup>d</sup>   | 0.46 <sup>c</sup>   | 0.56 <sup>b</sup>  | 0.65 <sup>a</sup>  | 0.11                         | XX  | NS |
| Faecal Saponin (g/d)   | 0.06                | 0.17                | 0.19                | 0.25               | 0.16               |                              |     |    |
| Digestible Saponin (%) | 75.00 <sup>a</sup>  | 58.58 <sup>b</sup>  | 58.88 <sup>b</sup>  | 58.33 <sup>b</sup> | 75.00 <sup>a</sup> | 9.84                         | NS  | X  |

<sup>b</sup>Probability for linear (L) and quadratic (Q) trends. \* $p < 0.05$ ; \*\* $p < 0.01$ ; \*\*\* $p < 0.001$ .

abcde: Means on same row with different superscripts differ =  $P < 0.05$

G0 = 0% *Gliricidia sepium* +100% *Panicum maximum* + Concentrate

G25 = 25% *Gliricidia sepium* +75% *Panicum maximum* + Concentrate

G50 = 50% *Gliricidia sepium* +50% *Panicum maximum* + Concentrate

G75 = 75% *Gliricidia sepium* +25% *Panicum maximum* + Concentrate

G100 = 100% *Gliricidia sepium* +0% *Panicum maximum* + Concentrate

*al.*, 1999). The phytate digestibility (%) was highest ( $P < 0.05$ ) in G0 (74.42) and lowest in G75 (57.35). showing a quadratic trend which also shows that there is little effect of level of inclusion in the degradability of the antinutritional factor but that the animals can degrade appreciable amount of this deleterious principle, This is in consonance with the findings of Aletor and Omodara (1994) and Oduguwa, *et al.*, (1998). There was complete degradation of the cyanide content in the feed, as supported by the findings of Rosenthal and Janzen (1979) who reported that the enzymes secreted by microbes in the rumen and large intestine have hydrolytic activity on cyanogenic glycoside to such an extent that there was little or none left in the resultant faeces. There was also a linear trend in the tannin digestibility (%) where highest (77.17) and lowest (60.70) was observed

in G100 and G75 respectively. Ruminants have rumen microbes that secrete enzymes tannase which degrade tannin especially if it is made of more hydrolysable than condensed. Apart from this mechanism, ruminants' saliva also possesses mucin that binds with tannin and release protein from the tannin-protein complex (Hoffman, 1987). Although there was a linear increase of intake of saponin with increase in legume level, the trend in digestibility was different and its linear trend not significant. There was appreciable but irregular trend in the degradability of saponin from the inclusion point of view. Saponin degradability had been confirmed by Rosenthal and Janzen (1979) who also attributed this to the inability of saponin to cause hypocholesteremia in ruminants. Apart from the abovementioned mechanisms for the attenuation of these antinutritional factors, herbivores have

some biochemical defence mechanisms against phyto-allelo chemicals and they include mixed function oxidase, epoxide hydrases, reductases, hydrolytic enzymes and group transfer enzymes. All these are intra-hepatic mechanism (Rosenthal and Janzen (1979). The various degradation by animals in the different diets without recourse or reference to the combinations or inclusion level has been reported by Devendra (1990) who observed that there are specific, breed and even individual differences in tolerance and utilization of plant allelo-chemicals or undesirable factors by ruminants. Other measures that can be embarked upon to attenuate or control or reduce these metabolites could be from two entities involved viz Plant and animal (ruminants). Plant selection and breeding of plants (spp, variety and accessions) with inherently low anti-nutrients factors can be employed (Devendra, 1990). In the same vein, molecular manipulation of the gene controlling these factors can be tried but the fear of tampering adversely with genes that are responsible for desirable attributes can not be ruled out. Furthermore, adoption of strategies like dilution techniques, a simple approach to

reduce toxicity is to feed the toxic plant in mixture with other plant, thus diluting the effective level of each compound can be used. Also, wilting technique can be employed; enzymes capable of degrading specific secondary compounds often occur with the compound in different structures in the same plant cells and reaction occurs when cell membrane are disrupted. Cutting management that will ensure feeding between flushies when the allelo-chemicals are at their peak in concentration and lastly, fertilizer management that will alleviate situation of nutritional stress in plant which stimulate the biosynthesis and secretion of these phytoalexins (Lowry, 1989). On the parts of the animals (ruminants): intrafusion / inoculation of bacterial species that can degrade anti nutrient from host rumen into rumen of other ruminants where multiplication and subsequent detoxification will continue (Allison, *et al.*, 1992) or even the genetic manipulation of otherwise rumen microbes with less degrading potential into those that can secrete enzymes that can detoxify the undesirable factors in herbage have been tried successfully (Keith, 1995)

#### CONCLUSION

Since ruminants (goats) can effectively utilize forages despite the inherent antinutritional factors they contain, research effort must be

directed towards harnessing these animals and forages potentials.

#### REFERENCES

- AJAYI, D.A., ADENEYE, J.A. and AJAYI, F.T. (2005).** Intake and nutrient utilization of West African Dwarf Goats fed Mango (*Magnifera indica*), Ficus (*Ficus thionninghii*), Gliricidia (*Gliricidia sepium*) foliages and concentrates as supplement to basal diet of Guinea grass (*Panicum maximum*), *World Journal Agricultural Sciences* **1**(2):184 -189.
- ALLISON, M.J., MARYBERRY, W.R., MCSWEENEY C.S. and STAHL, D.A. (1992).** *Synergistes jonesii* gen. Nov., sp. Nov: A rumen bacterium that degrades toxic pyridine diols. *System Applied Microbiology*. **15**:522-529.
- ALETOR, V.A. and OMODARA, O.A. (1994).** Studies on some leguminous browse plants with particular reference to their proximate, mineral and some endogenous anti-

- nutritional constituents. *Animal Feed Science and Technology*, **40**:343-442.
- ANUGWA, F.O., OKWORIA, I. and EKWUNO, P.O. (2000).** Feed intake, nutrient digestibility and nutrient value for goats of *Panicum maximum* and selected browse in the southern guinea savannah zone in Nigeria. In *Animal Production in the new millennium: Challenges and options. Proceedings of 25th Annual Conference of National Society for Animal Production at Michael Okpara University of Agriculture Umudike, Abia state. Nigeria* Pg 63-66.
- BEUTLER, H. BECKER, J., MICHAEL, G. and WALTER, E. (1980).** "Rapid method for determination of Oxalate". *Fresenius Journal of Analytical Chemistry*, **30**(1):186-187.
- BUDITANGENDJAJA and WINAELIZABETH (1995).** Chemical Evaluation of shrub legumes *International Journal of Agricultural Research* **17**(3):47 – 58.
- CONN, E.E (1981).** The Biochemistry of plants Vol 7. Secondary plant products. A.P New York Pg 479-498.
- DEVENDRA, C. and BURNS, M. (1985).** Goat production in the tropics in the tropics. C.A.B London. Pg 130.
- DEVENDRA, C. (1990).** The use of shrubs and Tree fodder by ruminants In: Devendra, c. (eds) shrub and Tree fodder for farm animals IDRC, Ohawa, Canada. Pp 42-68.
- EECHOAT, W.A. and DEPAEPE, M. (1994).** Total Phosphorus, phytate phosphorus and phytate activity in plant feedstuffs. *Animal Feed Science and Technology* **47**:19 -29.
- ENIOLORUNDA, O.O., JINADU, A.O., OGUNGBESAN, A.M. and T.O. BAWALA (2008.).** Effects of combination levels of *Panicum maximum* and *Gliricidia sepium* on Nutrient Digestibility and Utilization by West African Dwarf Goats fed cassava offal based Concentrate. *Journal of Animal Research* **2**(5) 149 – 153.
- ESMINGER, M.E. and OLENTINE, JNR. C.G. (1980).** Feeds and Nutrition: complete Esminger Pub. Co. (1980) California. U.S.A Pg 12-14.
- EVERS, A.D., BLAKENEY, A.B. and O'BRIEN, L. (1999).** Cereal structure and composition. *Australia Journal Agricultural Research*, **50**:629-50.
- HAGERMAN, A.C. and BUTLER, I.G. (1983).** Precipitation methods for the qualitative determination of Taninns. *Journal of Agriculture Food and Chemistry*, **26**:806-812.
- HOFFMAN, R.R. (1987).** Morphological evolutionary adaptation of ruminant digestive system. In aspect of digestive physiology in ruminants. Dobson A. ed. Cornell University press New York. U.S.A. Pg 1-26.
- IFUT, O.J. (1992).** Body weight responses of W.A.D goats fed *Gliricidia sepium*, *Panicum maximum* and Cassava peels Proceedings of the Joint feed Resources Networks Workshop Held in Gaborone, Botswana, 4 – 8th March. 1992 Pg 23- 29
- KEITH G. (1995).** Engineering gut flora of ruminant livestock to reduce forage toxicity, progress and problems. *TIBTECH* **13**:418-421
- LOWRY, J.B. (1989).** Toxic factors and problems: Methods of alleviating them in animals. In shrubs and tree fodders for farm animals. Proceeding Of Work shop in Densparay. Indonesia 24-29 July, IDRC Ontara Canada, Devendra, C. (ed) Pg 76-88.
- MAGA, J.A. (1983).** Phytate: Its occurrence, chemistry, food interactions nutritional significance and analytical method. *Journal of Agriculture food and Chemistry* **30**:1-9.
- NORTON B.W. (1994).** Antinutritive and Toxic factors in forage Tree legumes. In Forage Tree legume in Tropical Agriculture. Center of Agriculture Bioscience International Wallingford U.K. Gutteridge, R.C and Shelton, H.M (eds). Pg 203-214.

- ODUGUWA, B.O. ONWUKA, C.F.I. and OLAJOBI, H.O. (1998).** Antinutritional factors in foliage of some leguminous trees and shrubs. In Animal Agriculture in West Africa: The sustainability question Proceeding of 25th Annual Conference of National Society for Animal Production and Inaugural Conference of the West African Society for Animal Production (WASAP) Gateway Hotel Abeokuta, Nigeria March 21-26 Sept. Oduguwa, O.O, Fanimo A.O and Osinowo, O.A (eds) Pg 399-340.
- OGUNGBESAN, A.M, OGUNYEMI, A.O. and OLATITEDE Y. (2005).** Some *Tephrosia bracteolata's* (Guill et perr) antinutrients factors response to time of planting in the potential and limitations to self sufficiency in livestock production in Nigeria. Proceeding of 10th Annual Conference of Animal Science Association of Nigeria at University of Ado-Ekiti, Nigeria Sept 12 - 15. Dairo, F.A.S, Fajemilehin, S.O.K, and Onibi, G.E. (Eds) pg 222 – 224.
- OGUNGBESAN, A.M., AKINBOYE, O.O., APATA S.E. and OLUSANYA, T.P (2006).** Antinutrients Digestibility in West African Dwarf (WAD) goats fed *Tephrosia bracteolata* (Guill et perr) based diets. IN New challenge in animal production: The way forward. Proc. of 11th Annual Conference of Animal Science Association of Nigeria at I.A.R. & T. Ibadan. 18-21 Sept. Raji A.M., Oluokun, J.A and Odukoya, S.O (eds) Pg 63 – 65.
- OLOGHOBO, A.D. (1989).** Minerals and Anti-nutritional contents of forage legumes consumed by goats in Nigeria. In Africa small ruminant research and development. *Proceeding of annual Conference at Bamenda, Cameroon 18-23 June Africa small ruminant network Pg 219-229.*
- ONWUKA, C.F.I (1992).** Tannin and Saponin contents of some tropical fed to goats. *Tropical Agriculture (Trinidad) 69(2):176-180.*
- POONAN, V.R. and HAHN, S.K. (1984).** An automated enzyme assay for determining cyanide content of Cassava and its product. *Journal of Science Food and Agriculture 35:426-436.*
- PRESTORN, T.R. and LENG, R.A. (1987).** Matching ruminant production systems with available resources in the tropics and sub-tropics. Penambul book Armidale. Pg 24-25.
- ROSENTHAL, G.A. and JANZEN, D.H (1979).** Herbivores: Their interaction with secondary plant metabolites. Academic press (AP) Pg 4 - 25.
- S.A.S. (2002).** Statistical analysis system, user's guide statistical analysis institute Inc. Cary, North Carolina.
- SMITH, J.W., LARBI, A., JABBAR, M.A and AKINLADE, J. (1995).** Voluntary Intake by Sheep and Goats of *Gliricidia sepium* fed in three states and at three level of supplementation to a basal diet of *Panicum maximum*. *Agroforestry Systems 32:287-295.*
- STRONG, F.M. (1976).** Toxicants occurring naturally in foods. *International Journal of Food Science and Nutrition, 44:55.*
- SKERMAN, P. and RIVEROS, F. (1990).** Tropical Grasses. F.A.O Rome Pg, 80
- STRANGEL, P.J. (1993).** Nutrient cycling in sustaining crop. Livestock system in sub sahara: An overview in livestock and sustainable nutrient cycling in mixed farming system of sub-saharan Africa, Vol 2 Tech. papers of international conf. ILCA Addis Ababa Ethiopia, Nov 22 pg 38 -39.
- WILSON, T.R. (1995).** Livestock Production systems C.T.A Macmillan Wageningen, The Netherlands. Pg 10 – 11.